

GRANT NO: N00014-89-J-3070

Title: Characterization of Ground Squirrel Retinal Ganglion Cells

PROGRESS REPORT (Covers period 7-1-89 TO 12-31-89):PERSONNEL:

<u>Name</u>	<u>Title</u>	<u>Period of Service</u>		<u>% Effort</u>
		<u>From</u>	<u>To</u>	
N. Lugo-Garcia, Ph.D.	P.I.	7-1-89	12-31-89	25
R. E. Blanco, Ph.D.	Co-I.	7-1-89	12-31-89	20
Jacqueline Caban	Technician	7-1-89	12-31-89	100

Most of our efforts during the first six months of funding has been dedicated to our first specific objective: to characterize the dendritic arborizations of ganglion cells projecting to the dorsal lateral geniculate nucleus and the superior colliculus. Some technical difficulties were successfully dealt with and a few preliminary results have been obtained.

Rhodamine labeled latex microspheres were stereotaxically injected into both superior colliculi of three ground squirrels. Survival times ranged from ten days to two months. The long survival periods resulted in labeling of large numbers of retinal ganglion cells. These cells were often completely filled with the beads (Fig. 1). To visualize in detail the dendritic arborization of this population of ganglion cells, we made intracellular injections of Lucifer Yellow (Fig. 2). To date we have not received the micromanipulator and electrophysiological apparatus which is necessary to make these intracellular injections. We have been using similar, but not optimal, equipment at the Institute. We expect that once our equipment is complete and some remaining problems are resolved, the quality of our injections will improve, and we will be able to inject more cells in each preparation.

Other experiments in our laboratory relate to the nature of neurotransmitters/modulators in the ground squirrel retina (Lugo-Garcia, et al., 1990)*. This work will be presented at the American Association of Anatomists meeting to be held in Philadelphia on April, 1990.

*Lugo-Garcia, N., R.E. Blanco, T.E. Hughes and H. Karten, Localization of GAD-like and GABA-like immunoreactivity in the ground squirrel retina. Anatomical Record (1990) (submitted)

DISSEMINATION STATEMENT

Approved for public release
Distribution Unlimited

Approved for public release
Distribution Unlimited
All data and information
will be in black and
white

89 12 28 037

②
S
D
D
DEC 29 1989
FLECTE
DUC

LUGO-GARCIA, Nidza, Rosa Esther BLANCO*, Thomas E. HUGHES* and Harvey KARTEN, Dept. of Anatomy and Institute of Neurobiology, University of Puerto Rico, San Juan, Puerto Rico; Dept. of Neurosciences, UCSD, La Jolla, California. Localization of GAD-like and GABA-like immunoreactivity in the ground squirrel retina.

The identification of retinal neurotransmitters/modulators remains central to a complete understanding of the role of the retina in visual function. Immunohistochemical techniques have provided a precise means of identifying populations of retinal neurons which utilize specific neurotransmitters/modulators. We have used immunohistochemical methods to identify and characterize GABAergic neurons in the ground squirrel retina. Retinas were incubated with antibodies against glutamic acid decarboxylase (GAD) and gamma aminobutyric acid (GABA) and processed for fluorescence and/or avidin biotin labeling. Immunoreactivity was expressed in the inner nuclear layer (INL), inner plexiform layer (IPL), and ganglion cell layer (GCL). Immunoreactive neurons in the INL were identified as small amacrine cells with cell bodies near the inner nuclear/inner plexiform border. Labeled cells in the GCL may be either ganglion cells and/or displaced amacrine cells. The number of immunoreactive neurons was greater in retinal sections incubated with the GABA antiserum. Immunoreactive neurons in the INL and GCL gave rise to processes that entered the IPL. Immunoreactive processes ran through all IPL sublayers, but the staining intensity was highest in the innermost and outermost sublaminae. Thus, GAD and GABA immunoreactivity may be present in amacrine, displaced amacrine, and perhaps ganglion cells. (Supported by NIH Grant NS-07464, Navy Grant N00014-89-J-3070 and RCMC Grant RR-03051).

To assure good quality reproduction in the publication, the overall blackness and density of your typed abstract should match this paragraph.

A/V information:

Dual 35mm projection will be provided for platform presentations.

4' x 8' boards will be provided for poster sessions.

Special requests for additional AV equipment must be made in writing to Dr. Charles E. Slonecker, Program Sect., P.O. Box 1025, Ferndale, WA 98248 or FAX (604) 228-2316.

Preference for presentation:

- ☒ POSTER SESSION
☐ PLATFORM PAPER
☐ SPECIAL TOPIC SESSION

PREFERRED SUBJECT

CATEGORY (See "Notice of Annual Meeting")

1st CHOICE Neurosciences

2nd CHOICE Neurotransmitter Localization

FIRST AUTHOR STATUS

- ☒ Member of AAA
☐ Non-Member
☐ Post-Doctoral Fellow
☐ Student (Fill in section below)
☐ Other

Member Sponsoring Student Abstract

STUDENT FIRST AUTHOR

Full Name _____

Address _____

Phone _____

In experiments where patients or animals were used, were these experiments approved by a Human Ethics or Animal Care Committee? ☒ Yes ☐ No

Do not type in this space

CAUTION

Use this ORIGINAL FORM ONLY for preparation of Abstract. Use only this form for material applicable to the 1990 meeting.

Do not exceed limits of ruled box.

Do not draw the limit rules on copy intended for publication. The original reproduction form is printed in special non-reproducing blue ink to facilitate offset camera work. The use of other materials such as machine copies, rules drawn in ink or pencil, typewriter correction tape, china white, or any strike-on technique for corrections defeats this purpose. The sensitive offset camera "reads" all but the special form.

In the event corrections are necessary, the following technique is suggested:

1. Using a razor, carefully cut out the line containing errors. Do not cut into adjacent lines.
2. Retype the corrected line on white paper.
3. Place the retyped line under the Abstract form so that it shows through the "window" and tape it to the back of the form.